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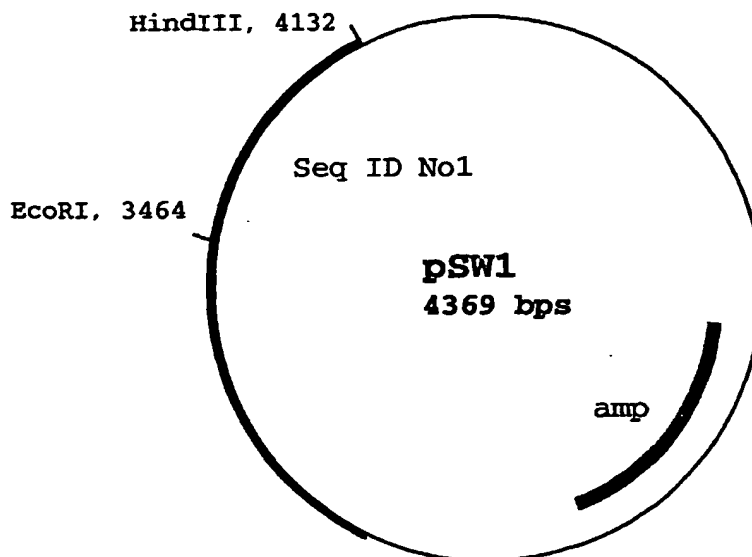
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(54) Title: VECTOR FOR INTEGRATION OF HETEROLOGOUS SEQUENCES INTO POXVIRAL GENOMES



(57) Abstract: The present invention provides a DNA vector comprising a nucleic acid sequence useful for inserting heterologous sequences into the genome of poxviruses by homologous recombination. The present invention relates also, *inter alia*, to recombinant poxviruses carrying heterologous coding sequences transferred by the vector according to the present invention.

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VECTOR FOR INTEGRATION OF HETEROLOGOUS SEQUENCES INTO POXVIRAL GENOMES

5 The present invention provides a new DNA vector comprising a nucleic acid sequence useful for inserting heterologous sequences into the genome of poxviruses by homologous recombination. The present invention relates also, *inter alia*, to recombinant poxviruses carrying heterologous coding sequences transferred by the vector according to the present invention.

10

Background of the invention

15 The successful worldwide eradication of smallpox via vaccination with live Orthopoxvirus, such as Vaccinia virus strain Western Reserve, Copenhagen or Ankara, stimulated in the early 80' research to study poxviruses in closer detail. Subsequently, said poxviruses were developed to well understood and easy-to-handle virus vectors or research tools, respectively (Moss, 1996). Today poxvirus vectors are used in various fields e.g. as expression vector or for the development of vaccines and therapeutic substances. The
20 main reasons for the high acceptance of poxvirus vectors are the following promising features: Firstly, the vector viruses are easy to manipulate, are highly stable and cheap to manufacture. Secondly, said vector virus can accommodate large amounts of heterologous DNA and proofed to be a versatile expression vector. Thirdly, said vector virus is easily administered
25 *in vivo* and succeeded in stimulating humoral and cellular immune responses. Accordingly, its use as a recombinant vaccine for protective immunisation against infectious disease or cancer made poxvirus vectors particularly attractive. Especially, Vaccinia virus, the best-known member of the Orthopoxvirus family, has been successfully used as recombinant
30 vaccine to protect against diseases in a large variety of animal models (Carroll et al., 1997; Sutter et al., 1994a).

To develop and establish recombinant vaccinia viruses several insertion sites have been used. The most prominent insertion site of the vaccinia genome is the locus of the viral thymidine-kinase (tk) gene (Mackett *et al.*, 1982). However, also other non-essential genes, such as the viral hemagglutinin and ribonucleotide reductase genes (Shida *et al.* 1987, Howley *et al.* 1996) or the naturally occurring deletion site II or III have been used to insert heterologous DNA sequences into the genome of vaccinia virus (Sutter *et al.*, 1994a). Construction of recombinant vector viruses carrying several heterologous genes or several immunogenic epitopes becomes more and more of general interest. Accordingly, there is a high need to identify further sites in the virus genome, which are suitable to insert further heterologous DNA sequences.

Insertion of heterologous DNA sequences into a poxviral genome bears the risk to destroy regions essential for the virus propagation due to a lack of complete understanding of the poxviral lifecycle. Although the sequence information of several poxvirus genomes (Goebel *et al.* 1990; Antoine *et al.* 1998) is available the function of most proteins encoded by the identified open reading frames is not known. Accordingly, it is still a complicated challenge to identify sites in the genome, which are suitable to stably take up heterologous DNA without destroying any sequences essential for viral replication and propagation.

Object of the invention

It is thus an object of the present invention to identify a new insertion site in the poxviruses genome and provide vectors suitable to direct the integration of heterologous DNA sequences into said insertion site.

Description of the invention

To achieve the foregoing and other objects, the present invention provides a vector comprising a nucleic acid sequence according to SeqID No. 1 or its
5 complementary strand. The nucleic acid sequence according to SeqID No. 1 is highly homologous with parts of the genomic sequences of a poxvirus genome. Due to this homology the nucleic acid sequence according to the present invention is capable to initiate homologous recombination between said sequence and the corresponding genomic sequences of
10 orthopoxviruses. Thus, the present invention provides a mean useful to direct integration of DNA sequences into the genome of different orthopoxviruses, preferably into the genome of modified vaccinia virus Ankara (MVA), but also of further related orthopoxviruses such as, e.g., Vaccinia virus strain Western Reserve or Copenhagen.

15 According to a preferred embodiment the nucleic acid sequence of the present invention is derived from modified vaccinia Ankara virus (MVA), especially from MVA, which has been isolated and deposited on January 27th, 1994 according to the Budapest Treaty at the European Collection of
20 Animal Cell Cultures (Salisbury, UK) under Deposit No.: V94012707.

The present invention further provides a vector comprising nucleic acid sequences, which hybridise under stringent conditions to the sequences according to SeqIDNo: 1 or its complementary strand. In the context of this
25 invention the term "vector" is understood as DNA vehicles of circular structure, such as plasmids, cosmids or artificial chromosomes. Said vector comprises in addition to the desired nucleic acid sequence regulatory sequences, selective marker genes and replicons enabling the autonomous replication of the vector. Hence, the vector according to the present
30 invention can easily be amplified in and isolated from unicellular host organism. Furthermore, the term "under stringent conditions" defines

parameters according to standard protocols (Sambrook et al., 1989), such as reaction temperature, formamide content or salt concentrations, which allow hybridisation of DNA-DNA sequences with a homology about and above 70%. As described above, also these sequences hybridising to the
5 corresponding sequence of the poxvirus genome and are thus, particularly, useful to integrate heterologous sequences into a genome of orthopoxviruses.

Additionally, the present invention provides a vector comprising
10 fragments of the above-mentioned nucleic acid sequence. These fragments comprise consecutive basepairs of said nucleic acid sequence and are also useful to integrate into the poxviral genome by homologous recombination. The length of said fragments is variable and fragments with only 30 basepairs being homologous to corresponding parts of the poxvirus
15 genome are already sufficient to initiate recombination events. However, to increase the efficiency of homologous recombination between the poxvirus genome and the fragments as used in the present invention, said fragments are preferably about and above 200 basepairs in length, more preferably about and above 300 or 500 basepairs in length.

20 To initiate homologous recombination the vector according to the present invention and a wildtype poxvirus is introduced into a host cell. During replication of the poxvirus genome homologous recombination between the nucleic acid sequence inserted into the vector and the corresponding
25 sequences of the poxvirus genome occurs. Since homologous recombination events occur only with a statistical probability of $1:10^3$ to $1:10^4$ any resulting recombinant poxvirus needs to be isolated. For this, e.g. a marker gene with functionally associated regulatory element is inserted into a cloning site of the nucleic acid sequence included in the vector. After
30 homologous recombination the resulting recombinant poxviruses are

isolated by screening for expression of said marker gene or by selection for the expression of a dominant-selection marker gene, respectively.

5 According to a further embodiment the vector of the present invention is particularly useful for insertion of a desired heterologous coding sequence into a poxviral genome. The term "heterologous" is used in the context of this invention for any combination of nucleic acid sequences that is not normally found intimately associated in nature. The heterologous genes according to the present invention are preferably selected from the group
10 of marker genes, therapeutic genes, such as anti-viral genes, anti-tumour genes, cytokine or chemokine genes, suicide genes, but also from host range genes or immunogenic epitopes. For insertion and/or expression of a desired heterologous coding sequences into a poxviral genome said heterologous coding sequence is inserted at a cloning site within the nucleic
15 acid sequence.

In general, a cloning site is a restriction enzyme recognition site. According to the present invention the preferred cloning site for the insertion of heterologous sequences is the restriction enzyme recognition site of the
20 EcoRI enzyme. This EcoRI site is unique in the nucleic acid sequence of the present invention, and is located between the two ORFs included in said nucleic acid sequence (Figure 1). Beside this, any further restriction enzyme recognition site, which is located in the non-coding regions between said two ORFs can be used as cloning site. Surprisingly, also any cloning site
25 located in one of the ORFs can be used for the insertion of heterologous sequences. Particularly, the inventors found that the destruction of said ORFs by such insertion into the ORFs does not hamper the viral life cycle or replication efficiency, respectively. Additionally, it was found by the inventors that the use of fragments according to the invention, which are
30 incorporated in the vector to initiate homologous recombination, likewise did not interfere with viral propagation or replication efficiency.

Generally, heterologous sequences to be integrated into a viral genome by homologous recombination are flanked on both ends by sequences being homologous to corresponding sequences of the viral genome. However, the present invention also includes vectors wherein the heterologous sequence is only flanked on one side by the above-mentioned nucleic acid sequence. According to the present invention also vectors comprising only one fragment and a desired heterologous sequence are useful to insert said heterologous sequence into a poxvirus genome.

To guarantee expression of an inserted heterologous coding sequence at least one transcriptional control element is additionally inserted into the cloning sites. This transcriptional control element is in functional association with the heterologous coding sequence, thereby controlling and/or allowing its expression. According to a further preferred embodiment of the present invention the transcription control element is derived from a poxvirus and/or is a consensus sequence of a poxvirus derived transcription control element.

According to still a further embodiment of the present invention the vector comprises at least two recombinogenic sequences, which flank one or more heterologous coding sequences, particularly sequences encoding e.g. a marker or a host range gene, and/or the transcription control element(s) inserted into the cloning site. The term "recombinogenic sequences" describes nucleic acid sequences, which, due to their similar or nearly identical structure, are capable to delete any sequence between said recombinogenic sequences by intragenomic homologous recombination. Accordingly, the sequences flanked by said recombinogenic nucleic acid sequences are only transiently inserted into the viral genome and are, subsequently, completely deleted. This deletion of sequences flanked by recombinogenic sequences is of particular interest for the isolation of recombinant poxviruses, which should comprise only a heterologous

coding sequence encoding a therapeutic or immunogenic gene, but no further marker or host range gene. For this, the marker gene, the host range gene and/or eventually also the transcription control element(s), but not the desired heterologous coding sequence, e.g. a therapeutic gene or immunogenic epitopes, are flanked by such recombinogenic sequences. After isolation of the recombinant virus, which is performed under a selection pressure upon the marker gene or the host range gene, the selection pressure is removed and thus, intragenomic homologous recombination to delete the marker or host range gene is allowed.

The present invention, furthermore, provides a recombinant poxvirus comprising in its genome the nucleic acid sequence transferred by the vector according to the present invention. Most preferably, this recombinant poxvirus is a recombinant MVA virus.

A further embodiment of the invention provides a method of treatment and/or prevention of an infectious disease or proliferative disorder. Said method comprises infection - either *in vivo* or *in vitro* - of a target cell population with recombinant poxviruses according to the present invention. Alternatively, according to this method the target cells are transduced- either *in vivo* or *in vitro* - with the vector according to the present invention and are infected, simultaneously or with a timelag, with any orthopoxvirus, included the recombinant poxvirus of the present invention. In this case, the poxvirus provides the cell with the poxviral replication and transcription machinery. As a consequence, the desired heterologous coding sequence incorporated in the vector and controlled by a poxvirus-derived transcriptional control element is expressed in the target cell. Target cells, which have been transduced or infected *in vitro*, can then according to the method of the present invention be applicated to a living animal body, including a human.

The invention provides the vector, the recombinant poxvirus and/or the target cells of the present invention useful for the treatment and/or prevention of an infectious disease or proliferative disorder. Furthermore, the vector, the recombinant poxvirus and/or the target cells according to the present invention are used for the production of a pharmaceutical composition, especially a vaccine, which is useful for *in vivo* and *in vitro* gene delivery and/or vaccination of mammals including humans, as described above.

Summary of the invention

The present invention, *inter alia*, comprises the following alone or in combination:

A vector for insertion of heterologous coding sequences into a poxviral genome, said vector including a nucleic acid sequence comprising one or more elements selected from the group consisting of:

(a) the nucleic acid sequence according to SeqID No. 1 or its complementary strand;

(b) a nucleic acid sequence which hybridizes under stringent conditions to the sequences as defined in (a);

(c) a fragment comprising at least 30 consecutive basepairs of the nucleic acid sequences as defined in (a) or (b);

the vector as above wherein the nucleic acid sequence is derived from a modified vaccinia Ankara virus (MVA);

the vector as above wherein additionally at least one transcriptional control element is included into at least one cloning site of said nucleic acid sequence;

the vector as above wherein the transcriptional control element is derived from a poxvirus genome or is the consensus sequence of a poxvirus derived transcriptional control element;

5

the vector as above additionally comprising at least one heterologous coding sequence, said heterologous coding sequence functionally associated with the transcriptional control element as above;

10 the vector as above wherein the heterologous coding sequence is selected from the group of marker genes, therapeutic genes, host range genes and/or immunogenic epitopes;

the vector as above comprising a recombinogenic sequence, which flanks
15 one or more heterologous coding sequences encoding marker genes, host range genes and/or the transcriptional control element as above;

a recombinant poxvirus comprising in its genome the nucleic acid sequence transferred by the vector as above;

20

the recombinant poxvirus as above wherein the poxvirus is a modified vaccinia Ankara virus (MVA);

a method of introducing a heterologous sequence into poxvirus genome
25 comprising

- (a) transduction of a host cell with the vector as above
- (b) infection of said host cell with a poxvirus, and
- (c) isolation of recombinant poxviruses;

30 a method of treatment and/or prevention of an infectious disease or proliferative disorder of a living animal body, including a human,

comprising application to said living animal body the recombinant poxvirus as above, and/or the vector as above, or application of said vector with any other poxvirus;

5 the method as above wherein the recombinant poxvirus is derived from an orthopoxvirus;

a target cell comprising the recombinant poxvirus as above and/ or the vector as above;

0

the vector as above, the recombinant poxvirus as above and/or the target cell as above for the treatment and/or prevention of an infectious disease or proliferative disorder;

5 the use of the vector as above, the recombinant poxvirus as above and/or the target cell as above for the production of a medicament for the treatment and/or prevention of an infectious disease or proliferative disorder;

0 a pharmaceutical composition comprising the vector as above, the recombinant poxvirus as above and/or the target cell as above, and a pharmaceutical acceptable carrier and/or diluent;

15 a pharmaceutical composition comprising the vector as above, a poxvirus, except the recombinant poxvirus as above, and a pharmaceutical acceptable carrier and/or diluent.

The following example will further illustrate the present invention. It will be well understood by a person skilled in the art that the provided example in no way may be interpreted in a way that limits the applicability of the technology provided by the present invention to this example, and the invention is therefore to be limited only by the full scope of the appended claims.

Example 1

10

Construction of the insertion vector

To obtain sequences suitable for recombination into a poxviral genome, a DNA fragment derived from the modified vaccinia Ankara virus (deposited according to the Budapest Treaty under Deposit No.: V94012707 at the European Collection of Animal Cell Cultures in Salisbury, UK) was amplified by conventional PCR using the following oligonucleotide primers:

- A24R_1; 5' - CCGAAAGCTTAATGAACGCCAGAGG - 3', SeqID No.: 2;
A27L_1c; 5' - AGGCTCGAGTAAGAGCGGCTATGAT - 3', SeqID No.: 3.

The oligonucleotide primers comprise, close to the 5' end and marked by underlining, a recognition sequence for the restriction enzymes HindIII (SeqID No.: 2) or XhoI (SeqID No.: 3) for subcloning of the resulting amplification product into a cloning vector. Accordingly, the specifically amplified sequence (SeqID No.: 1), which has a molecular weight of 1.7 kb, was subcloned Sall/HindIII into a pUC19 cloning plasmid (GenBank Accession No.: X02514). The resulting plasmid was designated pSW1 (Figure 2).

30

The subcloned insert has been sequenced, and this sequence was compared to other known sequences from vaccinia virus strains Copenhagen, WR, and MVA. It was found that said sequence comprises parts of the sequence of the MVA-ATI region. The ATI gene of most orthopoxviruses form a
5 dense cytoplasmic matrix embedding mature virions, so called inclusion bodies, which can be visualized by light microscopic examination of infected cells. Proposed ATI function is to provide higher stability and prolonged dissemination of infectious virus particles in the general environment. Among the orthopoxviruses are several members including
10 ectromelia virus, cowpox virus and racoon poxvirus produce this typical inclusion protein with a size of 130 to 160 kDa. However, other members of the orthopox genus, as e.g. vaccinia virus Western Reserve (WR), vaccinia virus Copenhagen or MVA, form no such inclusion bodies. This is a result of sequence deletions or frame-shift mutations leading to loss of coding
15 sequence and resulting in a truncated ATI-homologue. For example vaccinia virus WR expresses a 94 kDa ATI-homologue, while MVA and vaccinia virus strain Copenhagen expresses no such ATI-homologue.

For the further construction of insertion vectors, a naturally occurring
20 recognition site of the restriction enzyme EcoRI was used to split the amplified sequence into two segments. These segments serve as flanking regions (flank1, flank2), that initiate homologous recombination with a poxvirus genome. In between these flanking regions, vaccinia virus promoter sequences - e.g. of the 7,5 promoter (7.5) and/or the synthetic
25 promoter (sP) - as well as multiple cloning sites for the insertion of operably heterologous genes have been inserted. The resulting plasmids are designated pSW-7.5-sP, pSW-7.5, pSW-sP.

Additionally an expression cassette comprising the vaccinia virus host range
30 gene, K1L, fused to the EGFP fusion gene (isolated from the plasmid pEGFP, Clontech, GenBank Accession #: U76561) and the naturally

occurring K1L promoter, was inserted between the flanking regions as described above. This expression cassette is especially helpful for efficient selection and isolation of recombinant viruses. The resulting plasmid is designated pSWk1lgfp.

5

Generation of recombinant virus

For the generation of recombinant viruses 6-well tissue culture plates with cell monolayers of about 80% confluence are used. For the generation of recombinant MVA permissive cells such as chicken embryo fibroblasts are used. For the generation of recombinant vaccinia viruses of the strain WR African Green Monkey (Vero) cells have been used.

Firstly, the cell culture medium is discarded and the cells overlaid with serum-free medium containing wild-type poxvirus at a multiplicity of infection (MOI) of 0.01 (e.g. an inoculum 5×10^3 IU (infectious units) in 1 ml medium for one well with 5×10^5 cells). This mixture is incubated for 1 hour at 37 °C in 5% CO₂-atmosphere. Then, the inoculum is removed and the cells are washed twice with 2 ml OptiMEM per well.

Subsequently, the cell monolayer is overlaid with Lipofectin/plasmid DNA-mix (total volume: 1 ml) prepared as described by the manufacturer (GIBCO BRL) and using 15 µg plasmid DNA, of the pSWk1lgfp. The mixture is incubated for 5-12 hours at 37 °C in 5% CO₂-atmosphere. Then, the Lipofectin/plasmid DNA-mix is removed and the cells overlaid with 1.5 ml fresh medium supplemented with 10% FCS.

At 48 hours after infection, cell monolayer is detached with a cell scraper and the cells and medium are transferred into 2 ml-microcentrifuge tubes. The transfection harvest is stored at -20 to -80 °C.

Upon transfection of the plasmid pSWk1lgfp into poxvirus infected cells, the host range gene K1L fused to the EGFP gene were precisely recombined

into the site of the poxvirus genome, which is homologous with the flanking regions in the vector plasmid.

To isolate recombinant MVA viruses from non-recombinant MVA viruses a
5 host range cell, rabbit kidney (RK)13, was infected with virus material
obtained from the transfection experiment as described above. Previous
work had shown that MVA infection of rabbit RK13 cells results in an early
block of viral replication characterised by impaired synthesis of
intermediate viral RNA and lacking replication viral DNA. However, this
10 non-productive MVA infection of RK13 cells could be overcome by
coexpression of the vaccinia virus host range gene K1L (Sutter et al., 1994b).
Accordingly, inoculation of virus material obtained from said transfection
experiments into RK13 cultures resulted in the highly selective growth only
of recombinant viruses, which coexpressed the K1L gene. After five
15 consecutive passages on RK13 cells grown in 6-well or 96-well tissue
culture plates the virus MVA-K1LGFP was isolated. The absence of non-
recombinant MVA was demonstrated by PCR.

20 Additionally, the expression of the fused EGFP gene allowed a direct
monitoring of infection with recombinant virus via GFP fluorescence. The
direct monitoring is also used to identify and then isolate recombinant
Vaccinia viruses of the strain WR, which are not sensitive to a K1L
selection.

References:

- Antoine, G. et al., 1998, The complete genomic sequence of the modified vaccinia Ankara strain: comparison with other orthopoxviruses. *Virology*,
5 May 10;244(2):365-96.
- Carroll, M.W. et al. 1997: Highly attenuated modified vaccinia virus Ankara (MVA) as an effective recombinant vector: A murine tumor model. *Vaccine*
15, 387.
- 10 Fenner, F. et al. 1988: Smallpox and its eradication. World Health Organisation, Geneva.
- Goebel, S.J. et al., 1990, The complete DNA sequence of vaccinia virus.
15 *Virology*, Nov;179(1):247-66, 517-63.
- Howley, P.M. et al., 1996, A vaccinia virus transfer vector using a GUS reporter gene inserted into the I4L locus. *Gene*, Jun 26;172(2):233-7.
- 20 Mackett, M. et al. 1982: Vaccinia virus: a selectable eukaryotic cloning and expression vector. *Proc. Natl. Acad. Sci. USA* 79, 7415.
- Moss, B. 1996: Genetically engineered poxviruses for recombinant gene expression, vaccination, and safety. *Proc. Natl. Acad. Sci. USA* 93, 11341.
- 25 Sambrook, et al., 1989, *Molecular Cloning: a laboratory manual*, Cold Spring Harbor Laboratory Press publication, New York.
- Shida, H. et al., 1987, Effect of the recombinant vaccinia viruses that express
30 HTLV-I envelope gene on HTLV-I infection. *EMBO J.* Nov;6(11):3379-84.

Sutter, G. et al. 1994 a: A recombinant vector derived from the host range-restricted and highly attenuated MVA strain of vaccinia virus stimulates protective immunity in mice to influenza virus. Vaccine 12, 1032.

- 5 Sutter, G. et al. 1994b: Stable expression of the vaccinia virus K1L gene in rabbit cells complements the host range defect of a vaccinia virus mutant. J. Virol. 68, 4109.

C l a i m s

1. A vector for insertion of heterologous coding
5 sequences into a poxviral genome, said vector including a
nucleic acid sequence comprising one or more elements
selected from the group consisting of:
 (a) the nucleic acid sequence according to SeqID No.
1 or its complementary strand;
10 (b) a nucleic acid sequence which hybridizes under
stringent conditions to the sequences as defined in (a);

 (c) a fragment comprising at least 30 consecutive
basepairs of the nucleic acid sequences as defined in (a)
15 or (b).
2. The vector according to claim 1 wherein the nucleic
acid sequence is derived from a modified vaccinia Ankara
virus (MVA).
20
3. The vector according to claim 1 or 2 wherein
additionally at least one transcriptional control element
is included into a cloning site of said nucleic acid
sequence.
25
4. The vector according to claim 3 wherein the
transcriptional control element is derived from a
poxvirus genome or is the consensus sequence of a
poxvirus derived transcriptional control element.
30
5. The vector according to anyone of the preceding claims
1 to 4 additionally comprising at least one heterologous
coding sequence, said heterologous coding sequence
functionally associated with the transcriptional control
35 element according to claim 3 or 4.

6. The vector according to claim 5 wherein the heterologous coding sequence is selected from the group of marker genes, therapeutic genes, host range genes
5 and/or immunogenic epitopes.

7. The vector according to anyone of the claims 3 to 6 comprising a recombinogenic sequence, which flanks one or more heterologous coding sequences encoding marker genes,
10 host range genes and/or the transcriptional control element according to claims 3 to 5.

8. A recombinant poxvirus comprising in its genome the nucleic acid sequence transferred by the vector according
15 to any of the claims 3 to 7.

9. The recombinant poxvirus according to claim 8 wherein the poxvirus is a modified vaccinia Ankara virus (MVA).

20 10. A method of introducing a heterologous sequence into poxvirus genome comprising

- (a) transduction of a host cell with the vector according to anyone of the preceding claims 3 to 7,
- (b) infection of said host cell with a poxvirus, and
25 (c) isolation of recombinant poxviruses.

11. A target cell comprising the recombinant poxvirus according to claims 8 or 9 and/ or the vector according to anyone of the preceding claims 1 to 7.
30

12. The vector according to any of the preceding claims 1 to 7, the recombinant poxvirus according to claim 8 or 9 and/or the target cell according to claim 11 for the treatment and/or prevention of an infectious disease or
35 proliferative disorder.

13. The use of the vector according to any of the preceding claims 1 to 7, the recombinant poxvirus according to claim 8 or 9 and/or the target cell according to claim 11 for the production of a medicament
5 for the treatment and/or prevention of an infectious disease or proliferative disorder.

14. A pharmaceutical composition comprising the vector according to anyone of the preceding claims 1 to 7, the
10 recombinant poxvirus according to claim 8 or 9 and/or the target cell according to claim 11, and a pharmaceutical acceptable carrier and/or diluent.

15. A pharmaceutical composition comprising the vector according to anyone of the preceding claims 1 to 7, any poxvirus, except the recombinant poxvirus according to claim 8 or 9, and a pharmaceutical acceptable carrier and/or diluent.

20 16. A method of treatment and/or prevention of an infectious disease or proliferative disorder of a living animal body, including a human, comprising application to said living animal body the recombinant poxvirus according to claims 8 or 9, and/or the vector according
25 to anyone of the preceding claims 1 to 7, or application of said vector included in any other recombinant poxvirus.

17. The method according to claim 16, wherein the
30 recombinant poxvirus is derived from an orthopoxvirus.

1/2

Fig. 1

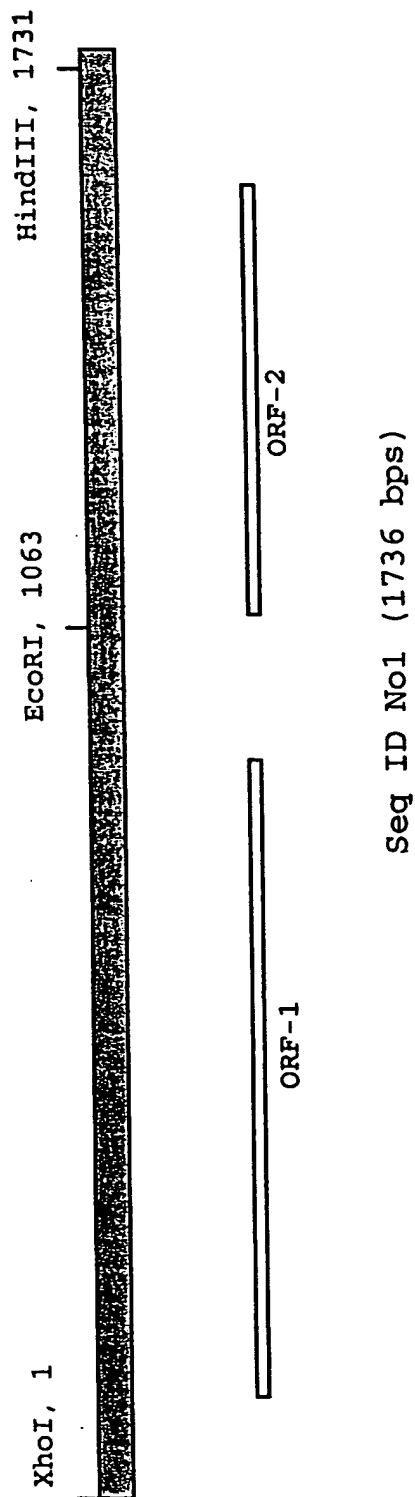
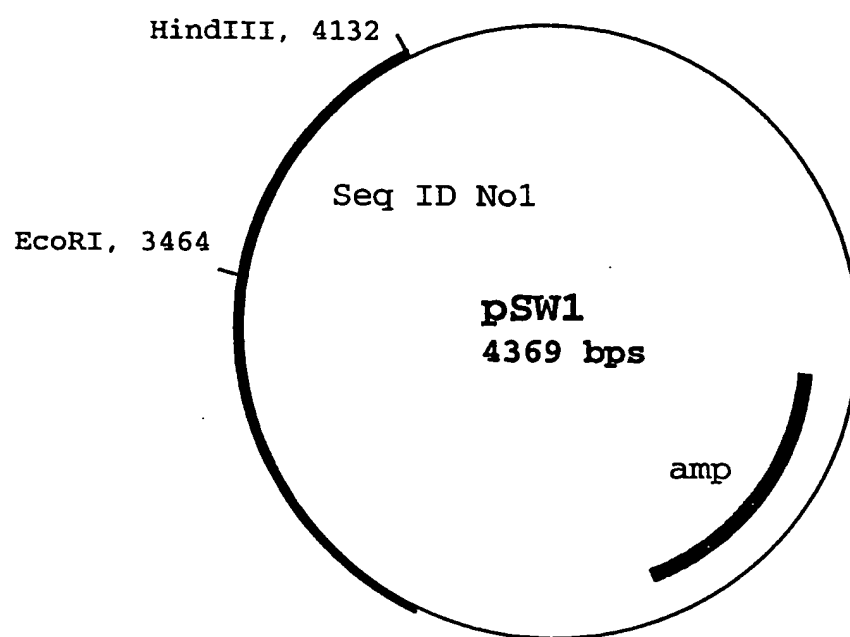


Fig. 2

<110> Gsf - Forschungszentrum für Umwelt und Gesundheit GmbH

<120> New vector for integration of heterologous sequences into poxviral genomes.

<130> BN31-Insertion site to MVA

<140>

<141>

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<170> PatentIn Ver. 2.1

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<213> Modified vaccinia Ankara virus

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gttgcgagca ttactgctt taaatctatg atagatgaaa catgggataa aaaaatcgaa 240
gcaaatacat gcatcagtag aaaacataga aacattattc acgaagttat tagggacttt 300
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<210> 3

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<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:
oligonucleotide primer

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INTERNATIONAL SEARCH REPORT

International publication No

PCT/EP 00/04786

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/86 C12N15/39 C07K14/07 A61K39/285

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

STRAND

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 261 925 A (TOA NENRYO KOGYO K.K. (JP); UNIV KYOTO (JP); SHIDA H.; FUNAHASHI S.) 30 March 1988 (1988-03-30) page 2, line 30-51 page 3, line 15,16; figure 3 page 3, line 41-44 page 7, line 3-11 page 7, line 55-61 page 8; claims	1-17
X	WO 97 02355 A (GSF-FORSCHUNGSZENTRUM UMWELT GESUNDHEIT GMBH (DE) SUTTER OHLMANN ERFLE) 23 January 1997 (1997-01-23) page 4, line 12-21 page 15; figures 1,4 page 28 -page 31; claims	15
A		2,9
	--- -/--	



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

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Date of the actual completion of the international search

24 October 2000

Date of mailing of the international search report

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INTERNATIONAL SEARCH REPORT

International Publication No

PCT/EP 00/04786

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	SUTTER G. AND MOSS B.: "Novel Vaccinia vector derived from the host range restricted and highly attenuated MVA strain of Vaccinia Virus" DEVELOPMENTS IN BIOLOGICAL STANDARDIZATION, vol. 84, 1995, pages 195-200, XP000611390	15
A	page 197; figure 2	2,9
X	WO 98 13500 A (BAVARIAN NORDIC RES INST (DK) MALAYSIA SARAWAK UNIV; GSF-FORSCHUNGSZTR) 2 April 1998 (1998-04-02)	15
A	page 7, paragraph 5	2,9
X	ANTOINE G. ET AL.: "Characterization of the Vaccinia MVA hemagglutinin gene locus and its evaluation as an insertion site for foreign genes" GENE, vol. 177, no. 1, 24 October 1996 (1996-10-24), pages 43-46, XP004043372 ISSN: 0378-1119	15
A	abstract	2,9
X	SCHEIFLINGER F. ET AL.: "Evaluation of the thymidine kinase (tk) locus as an insertion site in the highly attenuated Vaccinia MVA strain" ARCHIVES OF VIROLOGY, vol. 141, 1996, pages 663-669, XP000611391	15
A	abstract	2,9
X	US 5 185 146 A (HOFFMANN-LAROCHE INC. (US); ALTENBURGER WERNER) 9 February 1993 (1993-02-09)	15
A	abstract	2,9
X	US 5 443 964 A (DUKE UNIVERSITY (US); PICKUP DAVID J.; PATEL DHAVALKUMAR D.) 22 August 1995 (1995-08-22)	15
A	column 28, line 23-56	2,9
A	DATABASE WPI Section Ch, Week 198938 Derwent Publications Ltd., London, GB; Class B04, AN 1989-275691 XP002150903 & JP 01 202288 A (TOA NENRYO KOGYO KK), 15 August 1989 (1989-08-15) abstract	
	-/--	

INTERNATIONAL SEARCH REPORT

International Application No

PCT/EP 00/04786

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	JP 01 202288 A (UNIV KYOTO (JP); TOA NERYO KOGYO K.K. (JP)) 15 August 1989 (1989-08-15) figures 1-3 -----	
A	ANTOINE G. ET AL.: "The complete genomic sequence of the modified Vaccinia Ankara strain: comparison with other orthopoxviruses" VIROLOGY, vol. 244, 1998, pages 365-396, XP000887332 ISSN: 0042-6822 cited in the application ORF 137L page 377; table 1 page 384, left-hand column page 388, right-hand column -----	
A	MEYER H. ET AL.: "Mapping of deletions in the genome of the highly attenuated Vaccinia Virus MVA and their influence on virulence" JOURNAL OF GENERAL VIROLOGY, vol. 72, 1991, pages 1031-1038, XP000952390 abstract -----	

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/EP 00/04786

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP 0261925 A	30-03-1988	AU 614064 B	22-08-1991
		AU 7867787 A	23-06-1988
		CA 1322731 A	05-10-1993
		DE 3778809 A	11-06-1992
		JP 2063817 C	24-06-1996
		JP 7095956 B	18-10-1995
		JP 63192391 A	09-08-1988
		KR 9405590 B	21-06-1994
		US 5426051 A	20-06-1995
WO 9702355 A	23-01-1997	AU 721735 B	13-07-2000
		AU 6611096 A	05-02-1997
		BR 9609303 A	25-05-1999
		CA 2225278 A	23-01-1997
		CZ 9704241 A	18-03-1998
		EP 0836648 A	22-04-1998
		HU 9802217 A	28-01-1999
		JP 11509091 T	17-08-1999
		NO 980026 A	02-01-1998
		NZ 313597 A	28-01-1999
		PL 324347 A	25-05-1998
WO 9813500 A	02-04-1998	AU 4556597 A	17-04-1998
		EP 0951555 A	27-10-1999
US 5185146 A	09-02-1993	AT 97164 T	15-11-1993
		AU 2774389 A	20-07-1989
		DE 58906121 D	16-12-1993
		DK 11789 A	13-07-1989
		EP 0324350 A	19-07-1989
		ES 2059565 T	16-11-1994
		IE 63619 B	17-05-1995
		JP 2005860 A	10-01-1990
		JP 2589797 B	12-03-1997
		NZ 227562 A	28-04-1992
		PH 25747 A	18-10-1991
		ZA 8900097 A	27-09-1989
US 5443964 A	22-08-1995	US 5578468 A	26-11-1996
JP 1202288 A	15-08-1989	JP 2679711 B	19-11-1997
JP 01202288 A	15-08-1989	JP 2679711 B	19-11-1997